# OPTIMAL IMPERFECT PHYLOGENY RECONSTRUCTION AND HAPLOTYPING (IPPH) 

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#### Abstract

The production of large quantities of diploid genotype data has created a need for computational methods for largescale inference of haplotypes from genotypes. One promising approach to the problem has been to infer possible phylogenies explaining the observed genotypes in terms of putative descendants of some common ancestral haplotype. The first attempts at this problem proceeded on the restrictive assumption that observed sequences could be explained by a perfect phylogeny, in which each variant locus is presumed to have mutated exactly once over the sampled population's history. Recently, the perfect phylogeny model was relaxed and the problem of reconstructing an imperfect phylogeny (IPPH) from genotype data was considered. A polynomial time algorithm was developed for the case when a single site is allowed to mutate twice, but the general problem remained open. In this work, we solve the general IPPH problem and show for the first time that it is possible to infer optimal $q$-near-perfect phylogenies from diploid genotype data in polynomial time for any constant $q$, where $q$ is the number of "extra" mutations required in the phylogeny beyond what would be present in a perfect phylogeny. This work has application to the haplotype phasing problem as well as to various related problems in phylogenetic inference, analysis of sequence variability in populations, and association study design. Empirical studies on human data of known phase show this method to be competitive with the leading phasing methods and provide strong support for the value of continued research into algorithms for general phylogeny construction from diploid data.


## 1. INTRODUCTION

Sophisticated computational methods for data refinement and interpretation have become a core component of modern studies in human genetics. Computational methods have long been central to the study of phylogenetics, or evolutionary tree building, particularly the parsimony variants best suited to inferences of relationships over short time scales. The more specialized problem of haplotype reconstruction has also benefited tremendously from contributions from the fields of discrete algorithms and combinatorial optimization. In haplotype reconstruc-
tion, also called phasing, one seeks to separate the alleleic contributions of two chromosomes observed together in a diploid genotype assay. If we use the symbols 0 and 1 to represent homozygous and 2 to represent heterozygous alleles then ' 0221 ' is a genotype typed on four loci. Two pairs of haplotypes 0001,0111 and 0011,0101 are both consistent with the genotype and the goal of phasing is to correctly infer the true haplotypes, given the genotypes. The problem has relevance to basic research into population histories as well as to applied problems such as statistically linking haplotypes to disease phenotypes.

The field of computational haplotype reconstruction began with a fast and simple iterative method based on the idea that a haplotype that is observed in one individual is likely to be found in other individuals as well ${ }^{8}$. Various statistically motivated algorithms based on heuristic optimization techniques such as expectation maximization ${ }^{28}$ and Markov chain Monte Carlo methods ${ }^{35}$ have since been developed. Such algorithms provide significantly improved robustness and accuracy, although still poor scaling in problem size. Several combinatorial optimization methods have also been formulated based on Clark's original method ${ }^{20}$, although these have all been intractable in theory and practice. The advent of large-scale genotyping created a need for new methods designed to scale to chromosome-sized data sets.

Recently, a new avenue towards haplotype reconstruction was developed based on phylogenetic methods. In such an approach, one seeks to identify the ancestral history that could have produced a set of haplotype sequences from a common ancestor such that the inferred haplotypes could give rise to the observed genotypes. Gusfield ${ }^{19}$ showed that it is possible to directly and efficiently infer phylogenies that could explain an observed set of diploid genotypes provided the phylogenies are perfect, meaning that each character mutates only once from the common ancestor over the entire phylogeny. This problem is referred to as Perfect Phylogeny Haplotyping (PPH). Several subsequent results simplified and improved on this original method ${ }^{2,3,12,13}$. The work produced a fast, practical method for large-scale haplotyping, in which one breaks a large sequence into blocks consistent with perfect phylogenies and uses the phylogenies to phase those sequences.

The perfect phylogeny assumption is quite restrictive, though, and several approaches have been taken to adapt the perfect phylogeny method to more realistic models of molecular evolution. Data inconsistent with perfect phylogenies can arise from multiple mutations of a base over the history of a species or through the processes of recombination or gene conversion, which can assemble hybrid chromosomes in which different segments have different phylogenies. Heuristic methods have allowed some tolerance to recurrent mutations (for example,
the work by Halperin and Eskin ${ }^{23}$ ) resulting in the generation of imperfect phylogenies. Imperfect phylogeny haplotyping has proven to be very fast and competitive with the best prior methods in accuracy. Some recent work has been directed at provably optimal methods for imperfect phylogeny haplotyping (IPPH), resulting in a polynomial-time algorithm for the case when a single site is allowed to mutate twice (or a single recombination is present), under a practical assumption on the input data ${ }^{33}$. But the general IPPH problem remained open. It appears for the moment intractable in both theory and practice to infer recombinational histories in non-trivial cases.

Our Contributions: In this work, we solve the general IPPH problem and show for the first time that it is possible to infer imperfect phylogenies with any constant number $q$ of recurrent mutations in polynomial time in the number of sequences and number of sites typed. Our approach builds on both the prior theory on phylogeny construction from diploid data ${ }^{13,33}$ and a separate body of theory on the inference of imperfect phylogenies from haplotype data ${ }^{4}, 14,21,25,31$. Our algorithm reconstructs a $q$-near-perfect phylogeny in time $n m^{O(q)}$ where $m$ is the number of characters (variant sites typed), $n$ is the number of taxa (sequences examined), and $q$ is the imperfectness of the solution, defined below. The prior method of Song et al. ${ }^{33}$ that solves the 1-near-perfect phylogeny haplotyping problem relied on an empirically but not theoretically supported assumption that an embedded perfect phylogeny problem will have a unique solution. We relax this assumption to allow a polynomial number of such solutions and show that the relaxed assumption holds with high probability if the population obeys a common assumption of population genetics theory called Hardy-Weinberg equilibrium. We thus provide a theoretical basis for Song et al.'s empirical observation. We demonstrate and validate our algorithm by comparing with leading phasing methods using a collection of large-scale genotype data of known phase ${ }^{29}$. We find that our method is efficient and more accurate on blocks with small $q$. We further provide strong empirical support for the value of continuing research into accelerated algorithms for phylogeny construction from diploid data.

## 2. PRELIMINARIES

We begin by introducing definitions for parsimonybased phylogeny reconstruction. In such problems, we wish to study the relationship between a set of $n$ taxa, each of which is defined over a set of $m$ binary characters. Input $I$ will be represented by a matrix, where row set $R(I)$ represent taxa and column set $C(I)$ represent characters. In a haplotype matrix all taxa $r_{i} \in\{0,1\}^{m}$ and in a genotype matrix $r_{i} \in\{0,1,2\}^{m}$.

Definition 2.1. A phylogeny for $n \times m$ binary input matrix $I$ is a tree $T(V, E)$ and a label function $l: V(T) \rightarrow\{0,1\}^{m}$ with the following properties: $R(I) \subseteq l(V(T))$ and for all $(u, v) \in E(T)$, $d(l(u), l(v))=1$ where $d$ is the Hamming distance. That is, every input taxon appears in $T$ and the Hamming distance between adjacent vertices is 1 .

Definition 2.2. We define the following terms for a phylogeny $T$ on input $I$ :

- character $\mu(e)$ represents mutation on edge $e=(u, v)$ s.t. $l(u)[\mu(e)] \neq l(v)[\mu(e)]$.
- vertex $v \in V(T)$ is terminal if $l(v) \in R(I)$ and Steiner otherwise.
- length $(T)=|E(T)|$.
- phylogeny $T$ is optimal if length $(T)$ is minimized.
- penalty $(T)=$ length $(T)-m$.
- phylogeny $T$ is $q$-near-perfect if
penalty $(T)=q$ and perfect if penalty $(T)=0$.

We say that a character $i$ mutates on edge $e$ if $\mu(e)=i$. We will assume that both states 0,1 are present in all characters. Therefore the length of an optimum phylogeny is at least $m$. This provides a natural motivation for the penalty of a phylogeny as defined above. For simplicity, we will drop the label function $l(v)$ and use $v$ to refer to both a vertex and the taxon it represents.

The IPPH problem: The input to the problem is an $n \times m$ matrix $G$, where each row $g_{i} \in\{0,1,2\}^{m}$ represents a (genotype) taxon and each column represents a character. The output is a $2 n \times m$ matrix $H$ in which each row $h_{i} \in\{0,1\}^{m}$ represents a haplotype. Furthermore corresponding to every taxon
$g_{i} \in R(G)$, there are two taxa $h_{2 i-1}, h_{2 i} \in R(H)$ with the following properties:

- if $g_{i}[c] \neq 2$ then $h_{2 i-1}[c]=h_{2 i}[c]=g_{i}[c]$
- if $g_{i}[c]=2$ then $h_{2 i-1}[c] \neq h_{2 i}[c]$

The objective of the IPPH problem is to find an output matrix $H$ such that the length of the optimum phylogeny on $H$ is minimized. This problem is clearly NP-hard, since if matrix $G$ contains no 2 s , then the problem is equivalent to reconstructing the most parsimonious phylogenetic tree ${ }^{15}$. We therefore consider the following parameterized version of the problem. Given matrix $G$ and parameter $q$, we return matrix $H$ such that there exists an optimal phylogeny $T^{*}$ on $H$ with penalty $\left(T^{*}\right) \leq q$, under the assumption that such a matrix $H$ exists. Note that the PPH problem is a restriction of IPPH when $q=0$.

Definition 2.3. For a set of binary state taxa $S$ and a set of characters $C$, the set of gametes $G A M(S, C)$ is the projection of $S$ on characters $C$. In other words $\left(x_{1}, \ldots, x_{|C|}\right) \in G A M(S, C)$ i.f.f. there exists $s \in S$ with $s\left[c_{i}\right]=x_{i}$ for all $c_{i} \in C$. A set of characters $C$ shares $k$ gametes if $|G A M(S, C)|=k$. A pair of characters $i, j$ conflict if $|G A M(S,\{i, j\})|=4$.

Pre-processing: We perform a well established pre-processing step that ensures that for any optimal output matrix $H,(0,0) \in G A M(H,\{i, j\})$ for all $i, j \in C(H)$ (See the work of Eskin et al. ${ }^{13}$ ). We assume that the input matrix has no duplicate rows, since such rows do not change the optimal solution. Note that such a matrix should have at most $q+1$ more taxa than characters, since otherwise, it does not have a solution to the IPPH problem.

## 3. ALGORITHM

At a high level, our algorithm has the same spirit as the algorithm of Song et al. ${ }^{33}$. The algorithm guesses characters that mutate more than once and removes them from the input matrix. It then solves the perfect phylogeny haplotyping problem on the remainder of the matrix. Finally, our algorithm adds the removed characters back and performs haplotyping on the full matrix. In this section, we will use the same assumption of Song et al. ${ }^{33}$ : for any subset of characters of the input matrix, if a solution to perfect phylogeny haplotyping (PPH) exists, then it is
function solveIPPH(input matrix $G$ )
(1) guess $Q=\left\{i \in C(G) \mid \exists e_{1}, e_{2} \in E\left(T^{*}\right), e_{1} \neq e_{2}, \mu\left(e_{1}\right)=\mu\left(e_{2}\right)=i\right\}$
(2) let $M$ be matrix $G$ restricted to characters $C(G) \backslash Q$
(3) let $M^{\prime}$ be the unique solution to perfect phylogeny haplotyping of $M$
(4) let $H^{\prime}$ be matrix $M^{\prime}$ defined on characters $C(G)$ s.t. $\forall h_{2 i-1}^{\prime}, h_{2 i}^{\prime} \in R\left(H^{\prime}\right)$ and corresponding taxa $g_{i} \in R(G) . \forall \hat{c} \in Q . h_{2 i-1}^{\prime}[\hat{c}]=h_{2 i}^{\prime}[\hat{c}]=g_{i}[\hat{c}]$
(5) guess $\kappa=\left\{i \in C(G)\left|\exists j \in C(G),\left|G A M\left(t\left(T^{*}\right),\{i, j\}\right)\right|=4\right\}\right.$
(6) guess $G A M\left(t\left(T^{*}\right), \kappa\right)$
(7) loop for $c \in C\left(H^{\prime}\right) \backslash \kappa$
(a) $H^{\prime}:=\operatorname{processMatrix}\left(H^{\prime}, c\right)$
(8) for all couples $h_{2 i-1}^{\prime}, h_{2 i}^{\prime}$ in $H^{\prime}$
(a) resolve state 2 on $h_{2 i-1}^{\prime}, h_{2 i}^{\prime}$ s.t. $G A M\left(\left\{h_{2 i-1}^{\prime}, h_{2 i}^{\prime}\right\}, \kappa\right) \subseteq G A M\left(t\left(T^{*}\right), \kappa\right)$
function processMatrix (matrix $H^{\prime}$, character $c$ )
(1) initialize the set $\Delta:=\{c\}$
(2) while $|\Delta|>0$ do
(a) extract a character $c$ from $\Delta$
(b) for all $(2, c)$-couples $h_{2 i-1}, h_{2 i}$
i. for all $\hat{c} \in Q$ with $h_{2 i-1}[\hat{c}]=2\left(=h_{2 i}[\hat{c}]\right)$
A. if $\left|I N D\left(H^{\prime},\{c, \hat{c}\}\right)\right|=3$ then $\mathcal{G}(c, \hat{c})=I N D\left(H^{\prime},\{c, \hat{c}\}\right)$
else guess three gametes $\mathcal{G}(c, \hat{c})$
B. resolve state 2 in $\hat{c}$ on $h_{2 i-1}, h_{2 i}$ based on $\mathcal{G}(c, \hat{c})$
C. $\Delta:=\Delta \cup\left\{c^{\prime} \notin \kappa \mid h_{2 i-1}\left[c^{\prime}\right] \neq h_{2 i}\left[c^{\prime}\right]\right\}$
i.e. add to $\Delta$ characters $c^{\prime} \notin \kappa$ for which the current couple is also a $\left(2, c^{\prime}\right)$-couple
(c) if $\exists \hat{c} \in Q$. s.t. $(c, \hat{c})$ conflict or
for the set $H 2$ of all $(2, c)$-couples $G A M(H 2, \kappa) \nsubseteq G A M\left(t\left(T^{*}\right), \kappa\right)$ then
return no solutions
(d) remove $c$ from $C\left(H^{\prime}\right)$
(3) return $H^{\prime}$

Fig. 1. Algorithm to solve IPPH
unique. In Section 4, we show that the number of PPH solutions is polynomial with high probability.

Throughout the paper, we fix an arbitrary optimal phylogeny $T^{*}$, which we will use as a reference for expository purposes. Let $t\left(T^{*}\right)$ be the set of all taxa (Steiner vertices included) in $T^{*}$. Let $Q$ be the set of characters that mutate more than once in $T^{*}$. Since $|Q| \leq q$, we can find $Q$ by brute force in time $O\left(\sum_{i=1}^{q}\binom{m}{i}\right)=O\left(q m^{q}\right)$.

After finding $Q$, we can remove the characters to obtain matrix $M$ with character set $C(G) \backslash Q$. We now use a prior method to solve the perfect phylogeny haplotyping ( PPH ) problem on $M$ in time $O(n m)^{12}$. Let $M^{\prime}$ be the unique solution to the

PPH problem. The solution matrix $M^{\prime}$ contains $2 n$ taxa and $m-|Q|$ characters. We can now add the characters $Q$ to matrix $M^{\prime}$ to obtain $H^{\prime}$. For all $j \in Q$ and $h_{2 i-1}^{\prime}, h_{2 i}^{\prime} \in H^{\prime}$ and $g_{i} \in G$, taxa $h_{2 i-1}^{\prime}[j]=h_{2 i}^{\prime}[j]=g_{i}[j]$. Note that matrix $H^{\prime}$ contains state 2 only on the characters $Q$ (see Fig 3(b) for an example).

Definition 3.1. A pair of taxa $h_{2 i-1}^{\prime}, h_{2 i}^{\prime} \in R\left(H^{\prime}\right)$ is defined as a couple. For any $c \in C\left(H^{\prime}\right)$, if $h_{2 i-1}^{\prime}[c]=$ $2\left(=h_{2 i}^{\prime}[c]\right)$ or $h_{2 i-1}^{\prime}[c] \neq h_{2 i}^{\prime}[c]$ then $h_{2 i-1}^{\prime}, h_{2 i}^{\prime}$ is a (2, c)-couple.

Note that if contains both 0 and 1 in a couple of $H^{\prime}$, then the couple contained state 2
at character $c$ in the input (before perfect phylogeny haplotyping). Let $\kappa=\{i \in C(G) \mid \exists j \in$ $\left.C(G),\left|G A M\left(t\left(T^{*}\right),\{i, j\}\right)\right|=4\right\}$ be the set of characters that conflict with some other character in $t\left(T^{*}\right)$. Note that $Q \subseteq \kappa$. To complete the description and analysis of the algorithm we borrow the following definition from prior work ${ }^{13}$.

Definition 3.2. We say that a (unordered) couple $r_{1}, r_{2}$ induces $(x, y)$ at characters $\left(c, c^{\prime}\right)$ if $r_{i}[c]=$ $x, r_{i}\left[c^{\prime}\right]=y$ or $r_{1}[c]=r_{2}[c]=2, r_{1}\left[c^{\prime}\right]=r_{2}\left[c^{\prime}\right]=y$ or $r_{1}[c]=r_{2}[c]=x, r_{1}\left[c^{\prime}\right]=r_{2}\left[c^{\prime}\right]=2$. We define $I N D\left(H^{\prime},\left\{c, c^{\prime}\right\}\right)$ to denote the set of gametes induced by the couples of $H^{\prime}$ at $c, c^{\prime}$.

The goal now is to convert the $\{0,1,2\}$ matrix $H^{\prime}$ to a $\{0,1\}$ matrix $H$ such that the following correctness conditions are satisfied:
(1) for every $(2, c)$-couple in $H^{\prime}$ one of the two taxa should contain state ' 1 ' and the other ' 0 ' on character $c$ in $H$
(2) $G A M(H, \kappa) \subseteq G A M\left(t\left(T^{*}\right), \kappa\right)$, i.e. the set of gametes on $\kappa$ in $H$ is a subset of the set of gametes on $\kappa$ in $T^{*}$.
(3) $\left(\left|G A M\left(H,\left\{c, c^{\prime}\right\}\right)\right|=4\right) \Longrightarrow c, c^{\prime} \in \kappa$, i.e. a pair of characters share four gametes in $H$ only if they are both in $\kappa$.

In matrix $H^{\prime}$, if a couple contains state 2 at character $c$, then replacing it with state 0 on one taxa and 1 on the other is informally referred to as a resolution. The algorithm to solve IPPH is summarized in Figure 1. We now go into the details of the steps. The following lemma shows that Steps 5 and 6 of function solveIPPH can be implemented efficiently:

Lemma 3.1. Sets $\kappa$ and $\operatorname{GAM}\left(t\left(T^{*}\right), \kappa\right)$ can be found in time $m^{2 q} q^{O(q)}+O(n m)$.

Proof. We can easily identify $\kappa$ by brute force in time $O\left(m^{|\kappa|}\right)$. Since we do not know $|\kappa|$, this step can take time $O\left(2^{m}\right)$. We can however do better by performing such an enumeration over phylogenies as illustrated in Figure 2. First we construct the unique perfect phylogeny $T$ for the matrix $H^{\prime}$ restricted to $C(G) \backslash Q$ which contains $m-|Q|+1$ vertices in time $O(n m)^{12}$. Note that contracting edges $e \in T^{*}$ with $\mu(e) \in Q$ results in tree $T$ (see Figure $2(\mathrm{~b})$ ). We will begin with $T$ and add the edges $e, \mu(e) \in Q$ to
obtain $T^{*}$ as follows. Since we know $Q$ already, we can identify the set of $|Q|+q$ edges (labels $\mu$ ) in time $O\left(\binom{|Q|+q}{q}\right)=O\left(\binom{2 q}{q}\right)=O\left(4^{q}\right)$. There are $m-|Q|+1$ locations to add an edge that mutates a character in $Q$. All possible edge assignments can be enumerated in time $O\left((m-|Q|+1)^{|Q|+q}\right)=O\left(m^{2 q}\right)$. Each enumeration assigns a set of edges (multi-set on characters) $Q_{v}$ to each vertex $v$ of $T$. We now enumerate all possible rooted trees $T_{v}$ with edge labels in $Q_{v}$ for all vertices $v \in T$ in time $O\left((|Q|+q)^{|Q|+q}\right)=O\left((2 q)^{2 q}\right)$. Since the mutations in $C(G) \backslash Q$ are already fixed by the perfect phylogeny, the states in all vertices on characters $C(G) \backslash Q$ are known. For every root of $T_{v}$, we guess the states in all $Q$ characters in time $O\left(2^{q^{2}}\right)$, which can be improved to $O\left(q^{q}\right)$ (since this is equivalent to enumerating all tree structures with edges mutating $Q$ ). Note that since we guessed the states at the root of $T_{v}$, we know the states on all characters for all vertices in $T_{v}$ (for all $v$ ). Further, for any two roots of $T_{v}, T_{v^{\prime}}$, the path that connects them is given by the path connecting $v, v^{\prime}$ in the perfect phylogeny $T$ (see Figure 2(c)). Therefore, we can identify the edges that lie between two mutations of a character in $Q$. We can now find $\kappa$ since for all $i \in \kappa \backslash Q$, there exists $j \in Q$ and $e_{1}, e_{2}, e_{3} \in T^{*}$ such that $\mu\left(e_{1}\right)=\mu\left(e_{3}\right)=j, \mu\left(e_{2}\right)=i$ and $e_{2}$ lies in the path connecting $e_{1}, e_{3}$.

Since we know states in all characters of $T_{v}$ and $T_{v}^{\prime}$, we know the states in all the characters for every vertex in the path connecting them as well. We now consider the set of all vertices $V_{\kappa}$ that lie in the path connecting two mutations of the same character in $T$. It is easy to see that $G A M\left(V_{\kappa}, \kappa\right)=G A M\left(t\left(T^{*}\right), \kappa\right)$. We can therefore identify $\kappa$ and $\operatorname{GAM}\left(t\left(T^{*}\right), \kappa\right)$ in time $m^{2 q} q^{O(q)}+O(n m)$.

Figure 3 illustrates how the algorithm performs haplotyping given $\kappa$ and $\operatorname{GAM}\left(t\left(T^{*}\right), \kappa\right)$. For reference Figures 3(a) and 3(b) represent input matrix $G$ and matrix $H^{\prime}$ as found in Step 4 of solveIPPH. Function solveIPPH selects $c=2$ (Step 7). Function processMatrix determines $\mathcal{G}(2,6)=$ $I N D\left(H^{\prime},\{2,6\}\right)=\{(0,0),(0,1),(1,1)\}$ and guesses $\mathcal{G}(2,7)=\{(0,0),(0,1),(1,1)\}$ (Step 2(b)iA). Based on these two sets of three gametes, the $(2,2)$ couples, rows 9 to 12 , are resolved (Step $2(\mathrm{~b}) \mathrm{iB}$ ), Fig 3(c). Character 2 is removed (Step 2d). No-


Fig. 2. Algorithm to determine $\kappa$ and $G A M\left(t\left(T^{*}\right), \kappa\right)$ efficiently. (a) optimal phylogeny $T^{*}$ with $Q=\{6,7\}, \kappa=\{4,5,6,7\}(\mathrm{b})$ perfect phylogeny $T$ on characters $C(G) \backslash Q(c)$ the four edges mutating characters 6 and 7 are assigned to three vertices of $T$; rooted trees $T_{v}$ are constructed on the assigned edges; states on the three roots $0000000,0001010,0000111$ determine edges $\{4,5\}$ that lie between two mutations of 6,7 . Filled vertices form $V_{\kappa}$.
tice that a character $c$ is removed only when all the $(2, c)$-couples are completely resolved (rows 9 to 12). Function solveIPPH then selects $c=$ 3 (Step 7), Fig 3(d). Function processMatrix guesses $\mathcal{G}(3,6)=\{(0,0),(1,0),(0,1)\}$ and determines $\mathcal{G}(3,7)=\operatorname{IND}\left(H^{\prime},\{3,7\}\right)=\{(0,0)$, $(0,1),(1,1)\}$ (Step $2(\mathrm{~b}) \mathrm{iA})$, and using them $(2,3)$ couple, rows 7 and 8, are resolved(Step $2(\mathrm{~b}) \mathrm{iB})$, Fig $3(\mathrm{e})$. Since the pair of rows $(7,8)$ is also a $(2,1)$-couple character 1 is added to $\Delta$ (Step 2(b)iC). Character 3 is removed (Step 2d) and $c=1$ is extracted from $\Delta$ (Step 2a), Fig 3(f). Since there are no ( 2,1 )-couples, character 1 is removed (Step 2d), Fig $3(\mathrm{~g})$. This exhausts all $c \in C\left(H^{\prime}\right) \backslash \kappa$. Function solveIPPH then resolves state 2 in the first couple resulting in 0000,0010 which are both in $G A M\left(t\left(T^{*}\right), \kappa\right)$ (Step 8a), Fig 3(h). Since the next couple has no state 2 , we resolve the third couple which results in 0101,0111 (Step 8a), completing the algorithm, Fig 3(i). We now prove the main lemma that bounds the running time of our algorithm.

Theorem 3.1. If penalty $\left(T^{*}\right) \leq q$, then the algorithm described in Figure 1 returns a solution matrix $H$ that obeys all correctness conditions in time $n m^{O(q)}$.

Proof. To prove this theorem, we first need three simple lemmas.

Lemma 3.2. If $c_{1} \in \kappa$ and $c_{2} \in C\left(H^{\prime}\right) \backslash \kappa$, then $c_{1}, c_{2}$ share exactly three gametes in $t\left(T^{*}\right)$.

Proof. Every pair of characters share at least three gametes in $T^{*}$. Characters, $c_{1}, c_{2}$ cannot share four gametes since $c_{2} \notin \kappa$.

Lemma 3.3. For a pair of characters $c \notin \kappa, \hat{c} \in$ $Q$, given a set of three gametes $\mathcal{G}(c, \hat{c})$, there exists a unique resolution of state 2 in character $\hat{c}$ for any $(2, c)$-couple s.t. for resulting matrix $H^{\prime}$, $G A M\left(H^{\prime},\{c, \hat{c}\}\right) \subseteq \mathcal{G}(c, \hat{c})$

Proof. Let $r_{2 i-1}, r_{2 i} \in R\left(H^{\prime}\right)$ be a $(2, c)$-couple. By definition, $\quad r_{2 i-1}[c] \neq r_{2 i}[c]$. If $r_{2 i-1}[\hat{c}]=$ $r_{2 i}[\hat{c}]=2$ then the resolution will either create $G A M\left(\left\{r_{2 i-1}, r_{2 i}\right\},\{c, \hat{c}\}\right)=\{(0,0),(1,1)\}$ or $\{(0,1),(1,0)\}$. Only one of the two can be contained in set $\mathcal{G}(c, \hat{c})$ established between $c$ and $\hat{c}$.

Lemma 3.4. In matrix $H^{\prime}$, for $c \notin \kappa, \hat{c} \in Q$ if every $(2, c)$-couple is in $\{0,1\}^{m}$ then $\operatorname{GAM}(H,\{c, \hat{c}\})$ is fixed where $H$ is any matrix obtained from $H^{\prime}$ by resolution of $2 s$.

Proof. For any couple in $H^{\prime}$ if $h_{2 i-1}^{\prime}[\hat{c}]=h_{2 i}^{\prime}[\hat{c}]=$ 2 then according to the condition of the lemma, $h_{2 i-1}^{\prime}[c]=h_{2 i-1}^{\prime}[c]=s$. Therefore, for any couple $h_{2 i-1}, h_{2 i}$ in $H$ obtained by resolving state 2 on $\hat{c}$ will have the property that $\operatorname{GAM}\left(\left\{h_{2 i-1}, h_{2 i}\right\},\{c, \hat{c}\}\right)=$ $\{(s, 0),(s, 1)\}$.

Corollary 3.1. Although $\hat{c}$ can contain state 2 in matrix $H^{\prime}$, Step $2 c$ of function processMatrix in Figure 1 can test if $c, \hat{c}$ are conflicting. Furthermore, if $c, \hat{c}$ do not conflict at Step 2c, then the final matrix obtained by the algorithm will not have a conflict between $c, \hat{c}$.

We now return to the proof of the main theorem. Step 1 requires at most $q\binom{m}{q}$ enumerations. Lemma 3.1 shows that given the correct $Q$, both $\kappa$ and $G A M\left(t\left(T^{*}\right), \kappa\right)$ can be found in time $m^{2 q} q^{O(q)}+$


Fig. 3. Given $Q=\{6,7\}, \kappa=\{4,5,6,7\}, G A M\left(t\left(T^{*}\right), \kappa\right)=\{0000,0010,0011,1011,1010,0111,0101\}$, the algorithm chooses $c=2,3,1$. Based on a set of three gametes, it resolves states 2 in $\hat{c} \in Q$, in all ( $2, c$ )-couples. Shaded regions represent deleted (or ignored) characters and couples completely resolved by the algorithm. After exhausting all characters $c \in C(G) \backslash \kappa$, the algorithm considers the remaining couples and iteratively resolves states $2 \mathrm{~s} . \mathrm{t}$. the couples are in $G A M\left(t\left(T^{*}\right), \kappa\right)$.
$O(n m)$. We now bound the run time for function processMatrix.

Lemma 3.5. Total run-time for all calls to function processMatrix is $O\left(2^{q} n m^{2}\right)$.

Proof. First notice that once a character $c$ is removed from $\Delta$ at Step 2 a of function processMatrix, it is never added back throughout the execution of the algorithm. This is because, function processMatrix resolves all the states 2 present in $(2, c)$-couples and subsequently the character is deleted (or ignored) for the rest of the algorithm in Step 2d. This bounds the number of times Step 2a is executed throughout the algorithm as $O(m)$. To prove the lemma, we now bound the time for executing Steps 2 a through 2 d as $O\left(2^{q} n m\right)$.

The number of $(2, c)$-couples is $O(n)$ and therefore Step 2b loops for $O(n)$ times. The cardinality of $Q$ is at most $q$ and therefore the loop in Step 2(b)i executes $O(|Q|)=O(q)$ times. The time bound for guessing the set of gametes $\mathcal{G}(c, \hat{c})$ shared between $c$ and $\hat{c}$ is the hardest to analyze.

Consider any one call to function processMatrix. Let $c_{i}$ 's represent the characters added into $\Delta$ during the execution of a call. We show that if $\mathcal{G}\left(c_{j}, \hat{c}\right)$ is guessed at Step $2(\mathrm{~b})$ iA then for all future $c_{k}$ encountered during the execution of Step 2(b)iA, $\left|I N D\left(H^{\prime},\left\{c_{k}, \hat{c}\right\}\right)\right|=3$. When the algorithm guessed $\mathcal{G}\left(c_{j}, \hat{c}\right)$, by definition of $\left(2, c_{j}\right)$ couples, $h_{2 i-1}\left[c_{j}\right] \neq h_{2 i}\left[c_{j}\right]$ and by the condition on Step 2(b)i, $h_{2 i-1}[\hat{c}]=h_{2 i}[\hat{c}]=2$. Also $h_{2 i-1}\left[c_{k}\right]=$ $h_{2 i}\left[c_{k}\right]=x$ since otherwise $\left|I N D\left(H^{\prime},\left\{c_{k}, \hat{c}\right\}\right)\right|=3$
( $c_{k}$ and $\hat{c}$ cannot be identical characters). Let $c_{l}$ be the character extracted from $\Delta$ and used in the loop of Step 2 b during which $c_{k}$ was added into $\Delta$. This implies the existence of a couple $h_{2 i-1}^{\prime}, h_{2 i}^{\prime}$ which is both a $\left(2, c_{l}\right)$-couple and a $\left(2, c_{k}\right)$-couple. However $h_{2 i-1}^{\prime}[\hat{c}]=h_{2 i}^{\prime}[\hat{c}]=y$. Again, otherwise $\left|I N D\left(H^{\prime},\left\{c_{k}, \hat{c}\right\}\right)\right|=3$. Therefore matrix $H^{\prime}$ induces gametes $(x, 0),(x, 1),(0, y),(1, y)$ on characters $\left(c_{k}, \hat{c}\right)$. For all values of $x, y \in\{0,1\}$, this results in three induced gametes. The above proof therefore shows that the if condition in Step 2(b)iA fails at most $q$ times for any function call to processMatrix. Therefore the probability of all guesses performed at Step 2(b)iA being correct for any single call to processMatrix is at least $2^{-q}$. Equivalently, we suffer a multiplicative factor of $2^{q}$ in the run-time because of this step.

The check performed at Step 2c takes time $O(n m)$. Assuming $q<m$, the total running time for all calls to processMatrix combined is $O\left(2^{q} \mathrm{~nm}^{2}\right) . \square$

Using Lemma 3.2, we know that $c$ shares exactly three gametes with all characters $\hat{c} \in Q$ in $t\left(T^{*}\right)$. For character $c$, Step 2(b)iA (guesses) iterates over the set of all possible gametes shared between $c, \hat{c}$. Given the set of three gametes, Lemma 3.3 shows that there is a unique resolution of states 2 in the (2, c)-couples and this is performed in Step 2(b)iB. Correctness condition 1 holds by the definition of resolution. Step 2c of function processMatrix checks for conditions 2 and 3 . Using Corollary 3.1, we know that $c$ and $\hat{c}$ cannot conflict because of the reso-
lution of any of the remaining 2 s (ensures correctness condition 3). Finally, there can be no 2 s in character $c$ (since $c \notin Q$ ) or in any of the $(2, c)$ couples since it was just resolved. Step 8 of function solveIPPH iterates $n$ times. At each iteration, it performs a brute-force step of computing all possible ways of resolving the 2 s . Since only the characters in $Q$ can contain state 2 at this point, Step 8 a takes $O\left(m 2^{q}\right)$ time. This step also checks if the resulting gametes on characters in $\kappa$ are in the predicted set $G A M\left(t\left(T^{*}\right), \kappa\right)$ (ensures correctness condition 3). This shows that any matrix found by the algorithm obeys the correctness conditions and the running time is $n m^{O(q)}$. Finally, we know that there exists a set of three gametes $\mathcal{G}(c, Q)$ (as defined by $\left.t\left(T^{*}\right)\right)$ s.t. resolving based on $\mathcal{G}$ will ensure that $c, \hat{c}$ do not conflict and $G A M\left(H^{\prime}, \kappa\right) \subseteq$ $G A M\left(t\left(T^{*}\right), \kappa\right)$. Using these two observations, we know that if penalty $\left(T^{*}\right) \leq q$, then the algorithm finds matrix $H^{\prime}$ that obeys the correctness conditions in the stated time.

We now prove the correctness of our algorithm:
Theorem 3.2. Any solution matrix $H$ obeying all the correctness conditions is optimal.

Proof. The proof is constructive and demonstrates the procedure to construct a $q$-near-perfect phylogeny for $H$. The phylogeny along with correctness condition 1 guarantees that the returned matrix $H$ is an optimal solution.

Matrix $H$ satisfies the following two properties: if a pair of characters $c, c^{\prime}$ conflict in $H$, then $c, c^{\prime} \in \kappa$ (third correctness condition); a $q$-near-perfect phylogeny can be constructed on $\operatorname{GAM}(H, \kappa)$ (since $G A M(H, \kappa) \subseteq G A M\left(t\left(T^{*}\right), \kappa\right)$, second correctness condition). It can be shown that a $q$-near-perfect phylogeny can be constructed for any matrix that satisfies the above two properties (see Section 7 of Gusfield and Bansal ${ }^{21}$ ). Such a phylogeny is obtained by constructing the $q$-near-perfect phylogeny on on $G A M(H, \kappa)$, contracting the phylogeny to a vertex and constructing a perfect phylogeny on the remaining characters.

Theorem 3.3. The algorithm of Figure 1 returns an optimal solution $H$ to the IPPH problem in time $n m^{O(q)}$.

Proof. The proof follows directly from Theorems 3.1 and 3.2.

## 4. SOLUTIONS TO PPH

We assumed in the preceding sections, following prior work ${ }^{33}$, that the perfect phylogeny stage of the algorithm will find a unique solution, but this assumption does not necessarily hold. To guarantee optimality, the algorithm would need to enumerate over all solutions to the PPH sub-problem, increasing run-time proportionally. Prior work showed that the number of PPH solutions is at most $2^{k}$, where $k$ is the number of characters of $G$ that do not contain the homozygous minor allele ${ }^{13,19}$. In the worst case, this could be as large as $m$ and therefore the number of PPH solutions can be $2^{m}$. This however should not occur in practice since the underlying population typically follows a random mating model.

Hardy-Weinberg equilibrium states that the two haplotypes of any given individual are selected independently of one another at random. For any fixed character, let $p$ be the minor allele frequency and $(1-p)$ be the major allele frequency. Consider $G$, an $n \times m$ input genotype matrix to the PPH problem. The probability under Hardy-Weinberg that none of the $n$ taxa contain the homozygous minor allele is $\left(1-p^{2}\right)^{n}$. This probability could be large for very small values of $p$.

It is reasonable to assume that the value of $p>c$ for some constant $c$ since otherwise SNPs cannot be detected. In this case, with high probability in $n$ (at least $\left.1-\left(1-c^{2}\right)^{n}\right)$, a specific character contains the homozygous minor allele. Therefore in expectation the number of characters without a homozygous minor allele is at most $m\left(1-c^{2}\right)^{n}$. This expectation, exponentially tends to zero with $n$.

We now consider a more general setting when the value of $p$ is assumed to be uniformly distributed in $[0,0.5]$. Now, the probability that a specific character does not contain the homozygous minor allele is:

$$
\begin{gathered}
2 \int_{0}^{0.5}\left(1-p^{2}\right)^{n} d p \\
=2 \int_{0}^{1 / \sqrt{n}}\left(1-p^{2}\right)^{n} d p+2 \int_{1 / \sqrt{n}}^{0.5}\left(1-p^{2}\right)^{n} d p
\end{gathered}
$$

$$
\begin{gathered}
\leq \frac{2}{\sqrt{n}}+2 \int_{1 / \sqrt{n}}^{0.5}\left(1-p^{2}\right)^{n} d p \\
\leq \frac{2}{\sqrt{n}}+2 \sum_{i=1}^{\infty} \int_{i / \sqrt{n}}^{(i+1) / \sqrt{n}}\left(1-p^{2}\right)^{n} d p \\
\leq \frac{2}{\sqrt{n}}+\frac{2}{\sqrt{n}} \sum_{i=1}^{\infty}\left(1-\frac{i^{2}}{n}\right)^{n} \\
\leq \frac{2}{\sqrt{n}}+\frac{2}{\sqrt{n}} \sum_{i=1}^{\infty} e^{-i^{2}} \leq \frac{4}{\sqrt{n}}
\end{gathered}
$$

Now, if $n=\Omega\left(m^{2}\right)$, then using Chernoff bounds we can show that with high probability the number of characters that lack a homozygous minor allele is bounded by $2 \log m$ and therefore the number of solutions to the PPH problem is bounded by $m^{2}$.

This discussion answers the question raised by Gusfield on the theoretical estimate for the number of PPH solutions to expect from a coalescent model ${ }^{19}$. Furthermore, since PPH is always performed on SNP blocks with low diversity, it is not unreasonable to assume $n \gg m$. Since the number of solutions returned by PPH is $O\left(m^{2}\right)$, the IPPH algorithm described above with high probability just suffers $O\left(m^{2}\right)$ overhead for finding the optimal extension of each PPH solution.

## 5. EMPIRICAL VALIDATION

We demonstrate and validate our algorithm by comparing with leading phasing methods using a collection of large-scale genotype data of known phase from a high resolution scan of human chromosome $21^{29}$. The study identified common single nucleotide polymorphisms (SNPs) and typed them on 20 sequences through a method that directly identifies haplotypes rather than genotypes. The SNPs were partitioned into 4135 haplotype blocks by the authors of that study using a diversity test. We extracted haplotypes from the 'haplotype pattern' file provided in the supplementary material, which identifies distinct haplotypes in each block and provides some inference of missing data when it can be done unambiguously. We ignored haplotypes which still had significant missing data, replacing them with haplotypes randomly selected from the multinomial
distribution of fully-resolved haplotypes in the corresponding block in order to maintain 20 chromosomes per block. The haplotype blocks were divided into four sets based on their imperfectness $(q=0,1,2,3+)$ using a prior method ${ }^{31}$. A large majority of blocks $(98 \%)$ are 0 , 1 , or 2 near-perfect. We do not solve optimally for the $2 \%$ with imperfectness 3 or greater because the run-time for optimal solutions would be prohibitive. Such blocks can be solved non-optimally in practice by subdividing into smaller blocks of lower imperfectness, but such data would not be comparable to those solved optimally and is therefore omitted from our analysis. We then randomly paired haplotypes to produce 10 diploid genotypes per block as our final test set. Figure 4 shows that the length of the blocks is related to $q$, the imperfectness. The tails are heavier for larger values of $q$ as expected. For instance, this shows that a significant fraction of the $q=2$ blocks have large number of characters when compared with $q=1,0$.


Fig. 4. Distribution of the fraction of blocks as a function of the number of characters in the block. Data for more than 50 characters not shown.

We compared our method with two popular phasing packages. We ran the haplotyper ${ }^{28}$ package, which uses an expectation-maximization heuristic, with 20 rounds (the recommended value). We also ran the $\mathrm{PHASE}^{35}$ package, which employs a Markov Chain Monte Carlo method. Although

|  |  |  | Error rate |  |  | Total Run Time(secs) |  |  |
| :---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: |
| Perfectness | \#Blocks | \#SNPs | PHASE | haplotyper | our alg | PHASE | haplotyper | our alg |
| 0 | 3497 | 20816 | 0.11 | 0.11 | 0.17 | 3521.33 | 337.18 | 17.21 |
| 1 | 461 | 4211 | 0.53 | 0.47 | 0.35 | 805.62 | 80.62 | 8.77 |
| 2 | 93 | 1266 | 0.83 | 0.68 | 0.55 | 268.02 | 59.28 | 1111.18 |

Fig. 5. Empirical results on Chromosome 21. Blocks with 0, 1, 2 near-perfectness (accounting for 4051 out of 4135 blocks) were analyzed separately using the three algorithms. Error rate is the number of switch errors divided by the number of blocks. Total run time is the sum over all blocks.

PHASE can make use of additional information such as SNP positions, we provided it only the genotypes as input.

For our own method, with any given $q$, we enumerated all possible phylogenies that are at most $q$-near-perfect. Note that this does not guarantee finding all possible output matrices that are consistent with a $q$-near perfect phylogeny. Where multiple solutions were obtained, we selected the one that minimized the entropy of the haplotype frequencies. For the PPH sub-problem we implemented a fast prior algorithm ${ }^{13}$. We note that for simplicity, we do not implement the algorithm exactly as described. Rather, function processMatrix does not add characters into $\Delta$ to be processed iteratively. The implementation however always returns a haplotype output matrix and we report the switch errors ${ }^{33}$ for the output.

Table of Figure 5 summarizes the test results ${ }^{\text {a }}$. All methods provide comparable accuracy when blocks are perfect. We attribute the slightly worse performance of our method on perfect input to the fact that we do not use any procedure to select a maximum-likelihood output matrix among all perfect matrices, as is done for prior perfect phylogeny methods ${ }^{13}$. All three methods degrade in accuracy with increase in imperfectness. Our method, however, scales much better with imperfectness than do the other two, clearly outperforming them on 1-near-perfect and 2-near-perfect inputs. Imperfectness can result from recurrent mutations, recombinations or incorrect SNP inferences and we attribute our method's superior performance on imperfect data to the fact that it explicitly handles one of these factors while the others suffer from all three. Our method is extremely fast for $q=0$, where it
reduces to the perfect phylogeny algorithm of Eskin et al. ${ }^{13}$ and also substantially outperforms the competing methods for $q=1$. Our method's run time rapidly increases with larger $q$, though, as expected from the theoretical bounds.

## 6. DISCUSSION AND CONCLUSIONS

We have developed a theory for reconstructing phylogenetic trees directly from genotypes that is optimal in the number of mutations. As an immediate application, we solve the general IPPH problem. We demonstrate practical results that show great promise in accurately inferring phase from real data sets. Our results suggest that imperfect phylogeny approaches can lead to significant improvements in accuracy over other leading phasing methods. Run time, while very fast for perfect and almost perfect data, remains an obstacle for even modest $q$; this observation suggests a need for further research in improving theoretical and practical bounds for general $q$. Our new method has several immediate applications in computational genetics:

- Phasing: At present, the method is competitive with the most widely used tools in accuracy and, with optimizations, should become competitive in run-time for larger $q$. Both PHASE and haplotyper return confidence scores on its results, which might allow a slower, high-accuracy method such as ours to function as a fall-back for regions that those methods cannot infer with confidence.
- Phylogeny Construction: Our run times are competitive with typical times to be expected from other optimal phylogeny reconstruction algorithms, even when the input consists of haplotypes. Our approach may thus be considered

[^0]preferable to the standard practice of inferring haplotypes then fitting them to phylogenies.

- Haplotype Blocks Inference: Our method could serve as an improved means of identifying recombination-free haplotype blocks for purposes of association study design by more accurately distinguishing recurrent mutation from recombination. The blocks might thus be useful in improving statistical power in haplotype-based association testing.

Future empirical studies, enabled by our method, will be needed to better establish the nature of imperfectness in real genotype data and the degree to which better handling of recurrent mutations will be of use in practice.

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[^0]:    ${ }^{\text {a }}$ Haplotyper crashes on 18 blocks which were ignored in the calculation of its accuracy.

